Application Note: 375

# Collisionally-Induced Dissociation of Methimazole and Acetylsalicylic Acid: Proposed Mechanisms of Ion Formation using Hydrogen/Deuterium Exchange

Amin Kamel and Kevin Colizza, Department of Exploratory Medicinal Sciences, Pfizer PGRD, Groton, CT USA Patrick Jeanville, Thermo Fisher Scientific, Miami, FL USA

# **Key Words**

- TSQ Quantum Ultra AM™
- ESI
- High-resolution Mass Measurements
- Ionization Mechanisms

### Introduction

Acetylsalicylic acid (Aspirin) belongs to a class of medications known as non-steroidal anti-inflammatory drugs (NSAIDs) and has been among the most popular medications for treating mild to moderate pain. It also reduces inflammation, clotting of blood, fever, redness, swelling and discomfort caused by medical disorders such as headaches, infections and arthritis. Methimazole is used to treat hyperthyroidism, a condition that occurs when the thyroid gland produces too much thyroid hormone. [2]

Mass spectrometry has played a significant role in the characterization and analysis of small molecules such as Methimazole and Aspirin.<sup>[3,4]</sup> The present study was undertaken to assess the structures and mechanisms of formation of the principal fragment ions in the ESI mass spectra of these two drugs. The CID mass spectra of Methimazole and Aspirin and their corresponding deuterated analogs have been studied in both the positive and negative ion modes.

Decomposition mechanisms are proposed for the principal fragment ions using H/D exchange and the combination of multiple-stage CID at low collision energy with high-resolution mass measurements. The mass spectra of Methimazole and Aspirin can serve as useful models for structural determination of chemically or biologically modified Methimazole/Aspirin or related compounds.

# **Goals**

Thermo Scientific TSQ Quantum mass spectrometer systems feature HyperQuad quadrupoles. The hyperbolic shape of the electrodes yield higher ion transmission, minimizing false positives, while maximizing sensitivity. This work demonstrates the utility of the Thermo Scientific TSQ Quantum Ultra AM for assessing the structures and mechanisms of formation of principal fragment ions in the high-resolution CID mass spectra of famotidine and azithromycin in both negative and positive ion modes.

### **Experimental Conditions**

### **Chemicals and Reagents**

Acetic acid and ammonium hydroxide were purchased from Sigma-Aldrich (St. Louis, MO). HPLC grade methanol was acquired from Burdick and Jackson (Muskegon, MI), and HPLC grade water was purchased from J.T. Baker (Phillipsburg, NJ). Methimazole and

Acetylsalicylic acid were provided by Pfizer Global Research and Development (Groton, CT). Ammonium-<sup>15</sup>N, d<sub>4</sub> deuteroxide solution, CD<sub>3</sub>OD, D<sub>2</sub>O and Acetic acid-d<sub>4</sub> were purchased Sigma-Aldrich (St. Louis, MO). All chemicals were used as received.

Sample preparation: Stock solutions of Methimazole and Aspirin were prepared in HPLC grade methanol at 1.0 mg/mL. The stock solutions were then diluted with mobile-phase from each solvent system to give a final concentration of 20 μg/mL. No further sample preparation was required.

Sample analysis: Infusion analyses were performed on a TSQ Quantum Ultra AM triple-quadrupole mass spectrometer (San Jose, CA). The solvent systems used in this study were: A). 50:50 HPLC grade methanol and 0.1 acetic acid, B). 50:50 HPLC grade methanol and 50 mM ammonium hydroxide, C). 50:50 HPLC grade methanol and 0.1% Acetic acid-d<sub>4</sub>, and D). 50:50 HPLC grade methanol and 50 mM Ammonium-<sup>15</sup>N, d<sub>4</sub> deuteroxide. All infusion studies were conducted using the instruments' integrated syringe pump at a flow rate of 2.0 μL/min for total of four minutes.

MS conditions: The TSQ Quantum Ultra AM was calibrated in normal and high resolution modes with a solution of 1,3,6-Polytyrosine. Accurate mass calibration of the instrument was performed with a 50 pmol/μL solution of ammoniated polyethylene glycols (PEGs).

# **TSQ Quantum Ultra AM Conditions**

Ionization mode and source: Positive and Negative ESI Electrospray voltage: (+) 3.5kV; (-) -2.5kV

Sheath gas: 1 Auxiliary gas: 0

Ion transfer tube

temperature: 270 °C

Ion Transfer Tube offset: 35 V

Tube lens offset: 77V

Collision energy: 25eV (Methimazole); 15eV

(Aspirin)

Collision pressure: 1.2 mTorr
Q1/Q3 resolution: 0.1 Da FWHM
Accurate mass mode: Internal
Micro scans: 2



#### Results

CID of protonated methimazole (Figure 1) exhibits primary dissociations via eliminations of HCN and H<sub>2</sub>S (Figure 2). Supplementary losses of CH<sub>3</sub>, S and C<sub>2</sub>H<sub>3</sub>N are minor dissociation pathways from [M+H]<sup>+</sup> ion. Proposed mechanistic decompositions from the [M+H]<sup>+</sup> ion are given in Figure 3 and corroborate well with the empirical data. The CID mass spectrum of [M-H]<sup>-</sup> ion (Figure 4) shows loss of methyl group as the initial disso-

ciation step, followed by the elimination of HCN (Figure 5). The fragment ion NCS<sup>-</sup> at *m*/*z* 58 is also observed.

For protonated aspirin (Figure 1), elimination of  $H_2O$  (Figure 6) and the acetyl group (CH<sub>2</sub>CO) were the major decomposition pathways (Figure 7). The CID mass spectrum of [M-H]<sup>-</sup> of aspirin (Figure 8) was much simpler and showed the loss of CH2CO at m/z 137 as the primary dissociation pathway (Figure 9). The ensuing elimination of  $CO_2$  from the fragment ion at m/z 137 was a minor dissociation pathway.

Figure 1: Structures of Methimazole and Acetylsalicylic acid (aspirin)

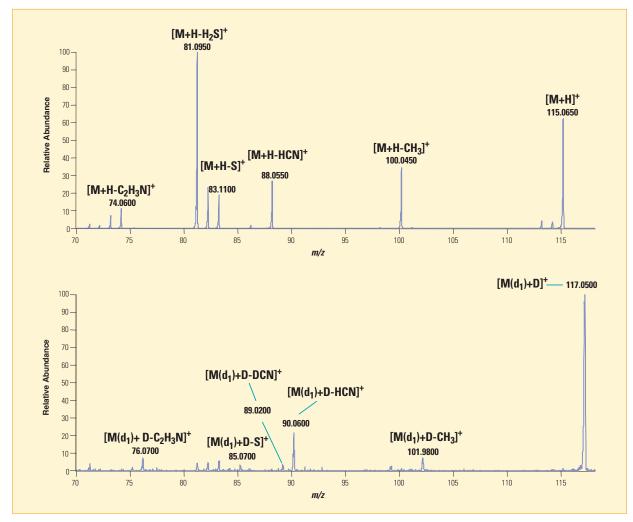


Figure 2: Positive ion ESI mass spectra of Methimazole (MW= 114): CID product ion spectra (MS/MS) of [M+H]\* at m/z 115 and the fully exchanged [M(d<sub>1</sub>)+D]\* at m/z 117. Deuteration was achieved by liquid phase H/D exchange method. MS and MS/MS experiments were performed on a TSQ Quantum Ultra AM mass spectrometer.

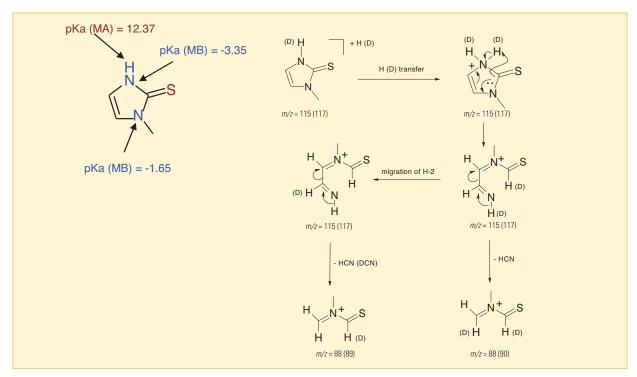


Figure 3: Proposed CID fragmentation mechanisms for the major fragment ions from protonated Methimazole at m/z 115 determined from H/D exchange patterns, high-resolution mass measurements and MS/MS experiments. Numbers in parentheses refer to deuterated fragment ions

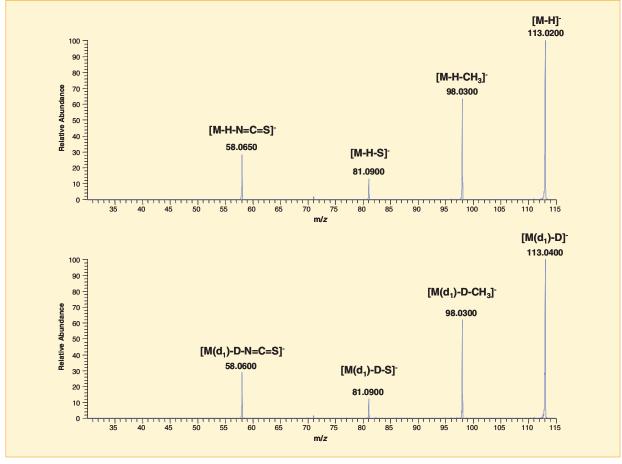


Figure 4: Negative ion ESI mass spectra of Methimazole (MW= 114): CID product ion spectra (MS/MS) of [M-H]<sup>-</sup> at m/z 113 and the fully exchanged [M(d1)-D]<sup>-</sup> at m/z 113. Deuteration was achieved by liquid phase H/D exchange method. MS and MS/MS experiments were performed on a TSQ Quantum Ultra AM mass spectrometer.

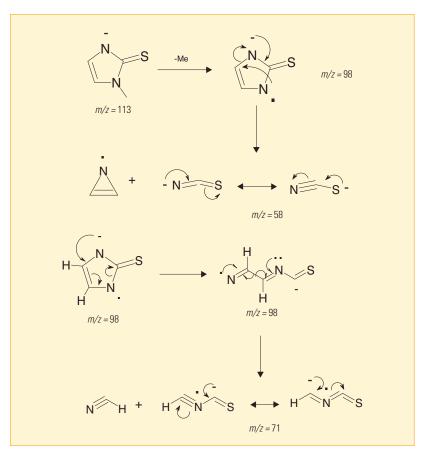


Figure 5: Proposed CID fragmentation mechanisms for the major fragment ions from deprotonated Methimazole at m/z 113 determined from H/D exchange patterns, high-resolution mass measurements and MS/MS experiments

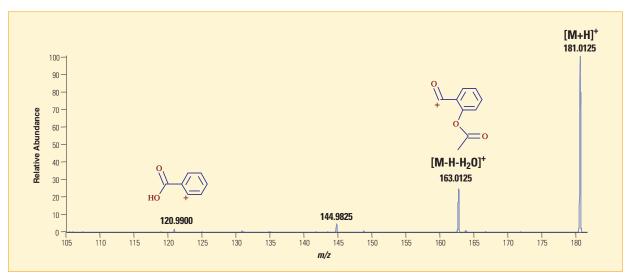


Figure 6: Positive ion ESI mass spectra of acetylsalicylic acid (MW= 180): CID product ion spectrum (MS/MS) of  $[M+H]^+$  at m/z 181. MS and MS/MS experiments were performed on a TSQ Quantum Ultra AM mass spectrometer

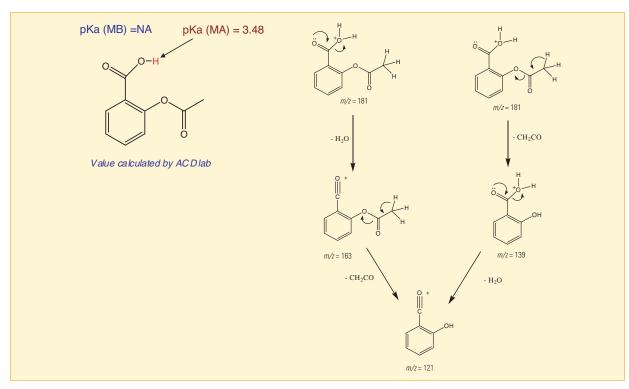


Figure 7: Proposed CID fragmentation mechanisms for the major fragment ions from protonated Acetylsalicylic Acid at m/z 181 determined from high-resolution mass measurements and MS/MS experiments

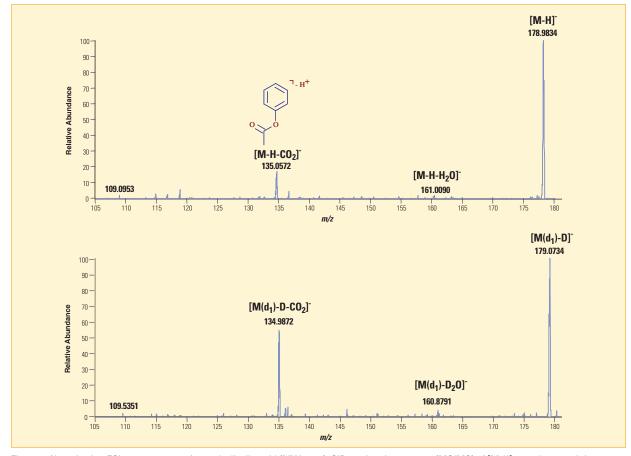


Figure 8: Negative ion ESI mass spectra of acetylsalicylic acid (MW= 180): CID product ion spectra (MS/MS) of [M-H] $^-$  at m/z 179 and the fully exchanged [M(d1)-D] $^-$  at m/z 179. Deuteration was achieved by liquid phase H/D exchange method. MS and MS/MS experiments were performed on a TSQ Quantum Ultra AM mass spectrometer.

$$m/z = 179$$
 $m/z = 137$ 
 $m/z = 93$ 

Figure 9: Proposed CID fragmentation mechanisms for the major fragment ions from deprotonated acetylsalicylic acid at m/z 179 determined from H/D exchange patterns, high-resolution mass measurements and MS/MS experiments

## **Conclusions**

Protonated methimazole dissociates primarily through elimination of HCN. Loss of CH<sub>3</sub>, S, and C<sub>2</sub>H<sub>3</sub>N are minor dissociation pathways from [M+H]+ ion. The CID mass spectrum of [M-H] ion show loss of methyl group as the initial dissociation step followed by the loss of HCN. The fragment ion NCS at m/z 58 is also observed.

For protonated aspirin, elimination of H<sub>2</sub>O and acetyl group (CH<sub>2</sub>CO) were the two major competitive decomposition pathways. The CID mass spectrum of the [M-H]- of aspirin was much simpler and showed the loss of CH<sub>2</sub>CO at m/z 137 as the primary dissociation pathway. Subsequent elimination of CO<sub>2</sub> from the fragment ion at m/z 137 was a minor fragmentation pathway.

The TSQ Quantum Ultra AM Mass Spectrometer system was designed to provide superior performance, while maintaining a level of flexibility not available on similar platforms.

- HyperQuad electrodes enabled acquisition of mass spectra at resolutions below 0.2 Da FWHM, without a significant reduction in signal response.
- High-resolution CID mass spectra were acquired using the TSQ Quantum Ultra AM mass spectrometer system of the fully-exchanged species. The high-resolution mass measurements aided greatly in structural assignments of fragment ions.
- The mass spectra of Methimazole and aspirin can serve as useful models for structure determination of chemically or biologically modified Methimazole and aspirin or related compounds.

#### References

- <sup>1</sup> Litovitz TL, Klein-Schwartz W, White S, Cobaugh DJ, Youniss J, Omslaer JC, Drab A, Benson BE (2001). "2000 Annual report of the American Association of Poison Control Centers Toxic Exposure Surveillance System." Am J Emerg Med 19 (5): 337-95.
- <sup>2</sup> Sgarbi, Jose A.; Villaca, Fabio G.; Garbeline, Benito; Villar, Heloisa E.; Romaldini, Joao H. Journal of Clinical Endocrinology and Metabolism (2003), 88(4), 1672-1677.
- <sup>3</sup> Mongillo, J.A.; Paul, J. Microchemical Journal (1997), 55(3), 296-307.
- 4 Reszka, Krzysztof J.; Wagner, Brett A.; Teesch, Lynn M.; Britigan, Bradley E.; Spitz, Douglas R.; Burns, C. Patrick. Cancer Research (2005), 65(14),
- <sup>5</sup> Kamel, A.M.; Brown, P.R.; Munson, B. Anal. Chem. 1999, 71(5), 968-977.
- <sup>6</sup> Kamel, A.M.; Brown, P.R.; Munson, B. Anal. Chem. 1999, 71(24), 5481-5492

In addition to these offices, Thermo Fisher Scientific maintains a network of representative organizations throughout the world.

**Africa** +43 1 333 5034 127

Australia +61 2 8844 9500

Austria

Belgium

+32 2 482 30 30

**Canada** +1 800 530 8447

**Denmark** +45 70 23 62 60

**Europe-Other** +43 1 333 5<u>034</u> 127

France

**Germany** +49 6103 408 1014

+39 02 950 591

Japan

+81 45 453 9100 **Latin America** 

+1 608 276 5659 Middle East

**Netherlands** 

South Africa

Spain

+34 914 845 965 Sweden/Norway/

Finland +46 8 556 468 00

**Switzerland** 

UK

+44 1442 233555 USA

+1 800 532 4752

www.thermo.com



an Jose, CA USA is ISO Certified

# Legal Notices

©2007 Thermo Fisher Scientific Inc. All rights reserved. All trademarks are the property of Thermo Fisher Scientific Inc. and its subsidiaries. This information is presented as an example of the capabilities of Thermo Fisher Scientific Inc. products. It is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others. Specifications, terms and pricing are subject to change. Not all products are available in all countries. Please consult your local sales representative for details.

View additional Thermo Scientific LC/MS application notes at: www.thermo.com/appnotes

