## **INSTRUCTIONS**



# Pierce<sup>®</sup> SDS-PAGE Sample Prep Kit

89888

Number

Description

**89888** SDS-PAGE Sample Prep Kit, sufficient materials to prepare 50 samples for SDS-PAGE

**Kit Contents:** 

**PAGE-prep Resin**, 1 ml, supplied as a DMSO-activated resin

**DMSO (dimethylsulfoxide)**, 27 ml **PAGE-prep Elution Buffer**, 5 ml

Spin Cups – Cellulose Acetate Filter, 50 each

Collection Tubes, 72 each

Lane Marker Non-reducing Sample Buffer (5X), 5 ml, contains 0.3 M Tris•HCl, 5% SDS, 50%

glycerol, lane marker tracking dye; pH 6.8

**Storage:** Upon receipt store kit at room temperature.

#### Introduction

For reliable SDS-PAGE analysis, protein samples must be prepared in a buffer free of interfering substances and at protein concentrations adequate for analysis. Many interfering substances are difficult to remove by traditional sample preparation methods. The SDS-PAGE Sample Prep Kit, however, provides a simple and effective method for concentrating samples while removing many chemicals, such as acids and bases, detergents, guanidine and ammonium sulfate, that interfere with SDS-PAGE analysis (Table 1, p.3). The proprietary PAGE-prep Resin binds protein in the presence of dimethylsulfoxide (DMSO). Interfering contaminants can then be removed by washing the protein-bound resin in an easy-to-use spin cup format. Protein elution is achieved in a buffer compatible with the BCA Protein Assay, allowing quantitation of a portion of the processed sample. The eluted proteins can then be combined with sample buffer and analyzed on the SDS-PAGE system of choice.

## **Important Product Information**

- This Sample Prep Kit is not recommended for purification or concentration of proteins in their active forms.
- Protein samples at pH 2-12 containing typical concentrations of salts (e.g., ammonium sulfate), organic solvents (e.g., acetonitrile), dyes (e.g., coomassie dye) and nonionic detergents can be effectively processed with the Sample Prep Kit.
- PAGE-Prep Resin will not effectively bind proteins in samples containing > 100 mM EDTA or > 4 M guanidine. For a list of known compatible substances, see the Substance Compatibility Table at the end of the protocol.
- Protein recovery is ~75-85% for most samples when using the recommended loading amounts (i.e., 1-70 μg of protein). After the sample has been processed, a portion of the recovered protein can be quantified using the BCA Protein Assay Kit (Product No. 23225).
- The PAGE-prep Resin cannot be regenerated and is intended for one-time use.



## **Additional Materials Required**

- Variable-speed bench-top microcentrifuge
- Water bath or heating block set to 60°C
- Reducing agent such as 2-Mercaptoethanol (Product No. 35602), TCEP (No. 77720) or DTT (No. 20290)

## Procedure for using the SDS-PAGE Sample Prep Kit

#### A. Resin Preparation and Sample Addition

- 1. Vortex the PAGE-prep Protein Binding Resin to evenly disperse the resin into a slurry.
- 2. Use a cut or large-orifice pipette tip to transfer 20 µl of resin slurry into the center of a spin cup.

**Note:** Resin particles may clog ordinary pipette tips.

3. Based on protein concentration, add 2-300 μl of sample containing 1-70 μg total protein to the resin. Cap tube and briefly vortex.

**Note:** Although 20  $\mu$ l of resin may bind as much as 130  $\mu$ g of protein, binding and recovery efficiency may be reduced with protein loads > 70  $\mu$ g.

4. Add a volume of 100% DMSO equal to the sample volume added in Step 3. Cap tube and briefly vortex.

Note: For best results, a minimal concentration of 50% DMSO must be maintained for optimal binding.

- 5. Incubate sample for 3-5 minutes at room temperature with occasional mixing to ensure maximum protein adsorption to the resin.
- 6. Centrifuge sample at  $2,000 \times g$  for 2 minutes. Discard flow-through and blot collection tube on a paper towel. Reinsert spin cup into the same collection tube.

#### B. Wash

- 1. Prepare Wash Solution by mixing 6 ml of DMSO with 6 ml of water, which makes sufficient solution for processing 20 samples. Store Wash Solution at room temperature.
- 2. Add 300 µl of Wash Solution to the resin. Cap tube and vortex until a homogeneous suspension is obtained
- 3. Centrifuge sample at  $2,000 \times g$  for 2 minutes. Discard flow-through and blot collection tube on a paper towel. Reinsert spin cup into the same collection tube.
- 4. Repeat wash one additional time for a total of two washes.

Note: If the original sample contained large amounts of detergent, an additional wash step may be required.

### C. Elution

- 5. Transfer the spin cup to a new collection tube and add 50 μl of Elution Buffer to the resin. Cap tube and briefly vortex to obtain a homogeneous suspension.
- 6. Incubate sample at 60°C for 5 minutes. After incubation briefly vortex the tube.
- 7. Centrifuge sample at  $2,000 \times g$  for 2 minutes. Discard spin cup and resin. Retain the collection tube containing eluate.

**Note:** If desired perform a second elution to recover additional protein; however, this is generally not required and may result in excessive sample dilution.

**Note:** Eluate may have a yellow tint; however, this will not affect downstream analysis.

- 8. (Optional) Assay a portion of the eluted sample for protein concentration using the BCA Protein Assay Kit.
- 9. Combine the appropriate sample volume for SDS-PAGE analysis with the supplied 5X sample buffer or with a sample buffer best suited for the specific electrophoresis system being used.
- 10. (Optional) Add reducing agent of choice (i.e., TCEP, DTT, or 2-mercaptoethanol).
- 11. Heat sample at 95°C for 5 minutes and proceed with SDS-PAGE according to manufacturer's recommendations.



**Table 1.** Concentrations of common SDS-PAGE interfering substances that are effectively removed by the SDS-PAGE Sample Prep Kit procedure.

Compatible			
Substance	Compatible Concentration*	Notes	
		Notes	
Acetonitrile	66%		
Ammonium sulfate	2 M		
Dyes	N/A	Resin discoloration may occur	
Strong chelating agents (EDTA)	100 mM		
Glycerol	25%		
Glycine pH 2.5	1 M		
Guanidine•HCI	4 M		
Imidazole	3 M		
Potassium thiocyanate	2 M		
Sodium chloride	1 M		
Reducing agents (DTT, TCEP, β-ME)	20 mM	Resin discoloration may occur	
SDS (Detergent)	20%		
Sucrose	40%		
Triton <sup>®</sup> -X 100, Tween <sup>®</sup> -20, CHAPs, OTG (Detergents)	10%		
Tris	1 M		
Urea	6 M		

<sup>\*</sup>Values represent concentrations of interfering substances that were successfully processed and do not necessarily represent the maximum.

## **Troubleshooting**

Problem	Possible Cause	Solution
Spin cup membrane clogs during processing	High protein concentration and/or detergents	Increase the centrifugation speed to $3,000 \times g$
Poor protein recovery	Poor binding efficiency	Increase DMSO concentration to > 60% in the binding solution, increase incubation time and further dilute potentially interfering substances
	Poor elution efficiency	Perform a second elution and combine sample
		Increase elution incubation time

#### **Related Pierce Products**

25200-25244 Precise™ Protein Gels (see catalog or web site for a complete listing)
24590 GelCode™ Blue Stain Reagent, sufficient reagent to stain 20 mini gels

24612 Pierce<sup>®</sup> Silver Stain Kit (SilverSNAP), sufficient reagents to stain 20 mini gels

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