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**Optimization of the Tango™ CCR6-*bla* U2OS Cell Line**

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**Tango™ CCR6-*bla* U2OS DA Assay Kit****Tango™ CCR6-*bla* U2OS cells**

Catalog Numbers – K1765 and K1761

**Cell Line Descriptions**

Tango™ CCR6-*bla* U2OS DA (Division Arrested) cells and Tango™ CCR6-*bla* U2OS cells contain the human Chemokine (C-C Motif) Receptor 6 (CCR6) linked to a TEV protease site and a Gal4-VP16 transcription factor stably integrated into the Tango™ GPCR-*bla* U2OS parental cell line. This parental cell line stably expresses a beta-arrestin/TEV protease fusion protein and the beta-lactamase reporter gene under the control of a UAS response element. Division Arrested (DA) cells are available as an Assay Kit, which includes cells and sufficient substrate to analyze 1 x 384-well plate.

DA cells are irreversibly division arrested using a low-dose treatment of Mitomycin-C, and have no apparent toxicity or change in cellular signal transduction. Both the Tango™ CCR6-*bla* U2OS cells and the Tango™ CCR6-*bla* U2OS DA cells have been functionally validated for Z' factor and EC<sub>50</sub> concentrations of MIP-3a (Figure 1). In addition, Tango™ CCR6-*bla* U2OS cells have been tested for assay performance under variable conditions.

## Validation Summary

Testing and validation of this assay was evaluated in a 384-well format using LiveBLazer™-FRET B/G Substrate.

### 1. MIP-3a dose response under optimized conditions

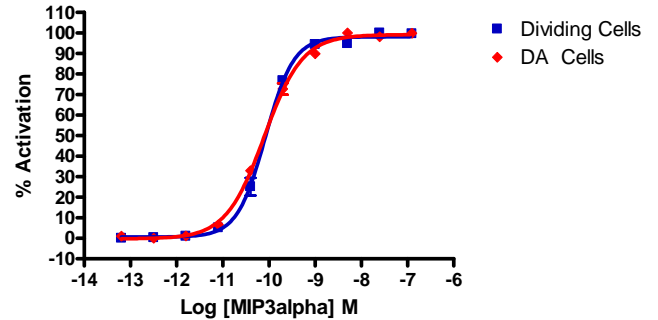
	DA Cells	Dividing Cells
EC <sub>50</sub>	0.079	0.084nM
Z'-factor	0.82	0.88
Recommended cell no. /well	= 10,000	= 10,000
Recommended Stim. Time	= 5 hrs	= 5 hrs
Max. [Stimulation]	= 125 nM	= 125 nM

### 2. Antagonist dose response

*No antagonists were commercially available at the time of publication of this document*

## Primary Agonist Dose Response

**Figure 1** — Tango™ CCR6-bla U2OS cells and Tango™ CCR6-bla U2OS DA cells dose response to MIP-3a under optimized conditions



Tango™ CCR6-bla U2OS cells and Tango™ CCR6-bla U2OS DA cells (10,000 cells/well) were plated in a 384-well format and incubated for 16-20 hours. Cells were stimulated with a dilution series of MIP-3a (Biosource (IVGN) PHC1234) in the presence of 0.1% DMSO for 5 hours. Cells were then loaded with LiveBLazer™-FRET B/G Substrate for 2 hours. Fluorescence emission values at 460 nm and 530 nm were obtained using a standard fluorescence plate reader and % Activation plotted for each replicate against the concentrations of MIP-3a.